CHEMISTRY OF β -TRIMETHYLSILYLETHANOL. II.¹ A NEW METHOD FOR PROTECTION OF AN ANOMERIC CENTER IN PYRANOSIDES^T

Bruce H. Lipshutz, $^{\star 2}$ Joseph J. Pegram and Matthew C. Morey Department of Chemistry University of California, Santa Harbara, CA 93106

 $SUMARY:$ Protection of the anomeric center in various carbohydrates as a β -trimethylsilylethyl glycoside is reported. The free sugar can be regenerated using $L\text{LBF}_{A}$ in acetonitrile.

Contemporary efforts in organic synthesis have now begun to focus heavily on the direct preparation of natural products in optically active form. One popular and very attractive means of achieving this goal is via transformation of inexpensive and readily available chiral educts³ such as carbohydrates.⁴ Synthetic manipulation of sugars must consider, at a very early stage, protection of the hemiacetal moiety. Traditionally, alkyl glycosides have been employed and conditions for subsequent unmasking generally fall into two categories: (1) acid hydrolysis; 5^{5} (2) reductive cleavage. 6 We now present an alternative procedure wherein novel 8-trinethylsilylethyl glycosides have been prepared which may ultunately be unraveled under the influence of fluoride ion to afford the corresponding free sugars.

The formation of β -trimethylsilylethyl glycosides was accomplished, in all cases studied, using β -trimethylsilylethanol (β -TMS ethanol)¹ and the appropriate sugar through four different routes (Methods A-D), the results of which are summarized in Table I.

Method A; Oxymercuration: 7 Treatment of either glucal (entries 1,2,6-8) or allal (entries 9,10) derivatives with Hg(OAc), (1 eq) / HOCH₂CH₂SiMe₂ (4 eq) in THF (1 M) at rt followed by ligand exchange (NaCl or NaOH) and reduction (NaBH_A) afforded the corresponding 2-deoxyglycosides in good yields.

Method B; Koenig-Knorr procedure: 8 Reaction of a glycosyl halide (an aminoglucosyl chloride, entry 3; a glucosyl bromide, entry 4; an arabinofuranosyl bromide, entry 11) with β -TMS

* Dedicated to Professor Harry H. Wasserman on the occasion of his sixtieth birthday.

ethanol (5 eq) in CHCl₃ (.5 M) containing CaSO₄ (2 eq), HgO (1 eq) and a trace of HgBr₂ at rt gave the desired saccharides.

Method C; Fisher Glycosidation: 9 Direct glycoside formation of mannose (entry 12) was realized using 10 eq 6-TMS ethanol in refluxing THF (16 M) containing a catalytic amount of $PCL₃$ or TsOH. Trituration of the concentrated reaction mixture with pentane permitted recovery of the majority of β -IMS ethanol.

Method D; Glycal Rearrangement: 10 Dissolution of tri-O-acetyl-D-glucal in dry benzene (1 M) at 25° C followed by addition of HOCH₂CH₂SiMe₃ (1.5 eq) and BF₃· Et₂0 (cat.) led to a 2,3 unsaturated glycal (entry 13).

It was expected from previous experience¹ that n-Bu_ANF would be the reagent of choice for deprotection owing to the structural similarities between SEM ethers and the β -TMS ethyl gly-

$\mathbb{C}^{\mathbb{C}}$) n $Me_3Si \sim 0$ or vs $Me_3Si \sim 0$ or α .

cosides. We ware, therefore, quite surprised to fmd that essentially all sources of fluoride examined (e.g., n-Bu₄NF, Et₄NF, CsF, HgF₂, KF, MgF₂, ZnF₂, PhCH₂NMe₃F, Me₄NF, pyr.HF) in a variety of solvents (THF, HMPA, DMSO, DMF, CH₃NO₂, CH₃ON) gave, in most cases, little or no reaction. A recent report, 11 however, describing the use of LiBF_A for the cleavage of silyl ethers has led to our effective use of this reagent for glycoside breakdown. Hence, exposure of a β -TMS ethyl glycoside to 5-10 eq of dry LiBF₄ in distilled (CaH₂) CH₃CN (\sim 1.5 M) at <u>ca</u>. 70° for 3-8 h gave good yields of free sugar. Simular attempts in CH₃NO₂, DMSO and DMF gave no reaction.

In order to briefly examine the mechanism of deprotection, the following additional observations have been made: (a) dry LiBF_A did not cleave a methyl glycoside, suggesting F^- was indeed involved; (b) both $NABF_A$ and $RBF_A/18$ -crown-6 ether were completely ineffective, although this is likely a reflection of their limited solubility in CH₃CN; (c) LiF/CH₃CN returned only starting material but addition of $BF_3F_3F_2O$ (LiF : $BF_3F_3F_4O = 1:1$) led to deprotection; (d) the β -anomer within a mixture appears to unravel at a far greater rate (< 1 h) relative to the α isomer; (e) ß-TMS ethyl furanosides (e.g., entry 11) apparently do not undergo cleavage under these or related conditions, returning only starting material.

From these and other points mentioned above, it is likely that a 'push-pull' mechanism is operating where boron derivatives act as both the source of fluoride 11 and Lewis acid through dissociation of LiBF₄. Since generation of some BF₃ seems necessary for complexation (or

> $\text{LiBF}_4 \div \text{Li}^+ + \text{BF}_4^ BF_4 \pm F + BF_3$

'pull'), LiBF₄ functions as a gradual release of both F⁻ and BF₃. CH₃CN, the only non-oxygencontaining solvent, possibly does not compete with the substrate for association. The nature of the complexation in the pyranoside versus furanoside systems, as well as the penchant for rapid cleavage of the 8-anomer remains unclear at this time.

^a Satisfactory ir, nmr, mass spec, as well as combustion analysis data were obtained for all new compounds bSee text ^cDetermined by either HPLC, NMR, or isolation ^dIsolated, chromatographically pure. ^e Free sugars were compared with authentic samples ^f See Ref 12 ⁹ Not attempted. ^h Both the β -TMS ethyl group and benzylidene acetal were cleaved ⁱ Not determined.

In conclusion, the methodology described herein represents a convenient and efficient means of protecting an anomeric center. A variety of carbohydrates may function as precursors of β -trimethyls1lylethyl glycosides, the subsequent unmasking of which can be carried out with a commercially available reagent (LiBF_{$_A$}). The anticipated cleavage of an anomeric center, protected in this fashion, with fluoride ion under aprotic and non-reductive conditions should expand the flexibility of carbohydrates in synthesis. Further studies on the useful applications of 8-TMS ethanol are in progress.

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References & Notes

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